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Review article

Port-site metastasis in laparoscopic gynecological oncology surgery: An overview



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ABSTRACT

Despite the low frequency, port-site metastases are associated with poor outcomes in patients and account for significant patient morbidity. They remain a challenging complication of laparoscopic procedures for gynecologic malignancies. A comprehensive, systematic search for published studies was conducted using the PubMed databases. Various mechanisms of port-site metastases are addressed in the relevant literature. The review of the articles points out that in the development of port-site metastases, the major role is played by biologically aggressive diseases, tumor manipulation, wound contamination, and surgery-related factors. The advantages of laparoscopic oncologic surgery are unquestionable. Further investigations of the mechanisms of port-site metastasis would contribute to the prevention of this insidious pathology.

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Introduction

Laparoscopic surgery has been used for treatment of oncological patients for the past 30 years. The first reports concerning laparoscopy implementation in oncogynecology date back to the 1970s.^{1–3} It is undebatable that laparoscopic surgery has numerous advantages. It affords a safe and less invasive modality for both diagnostic and major operative procedures.⁴ Its safety and feasibility have been proved by numerous authors, who published their experience with total laparoscopic radical hysterectomy.^{5–7}

Laparoscopy has several significant advantages in oncologic patients.^{8–14} Oncologic and immunologic functions are much better preserved after laparoscopic surgery.^{15–17} Additionally, specific to gynecologic malignancies, shorter intervals to postoperative treatments can also be listed as advantages to minimally invasive

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surgery.¹⁸ Patients who undergo minimally invasive surgery can often begin adjuvant therapy relatively quickly after their initial surgery because of the shorter recovery time.¹⁹

As main complications of laparoscopic oncology, the authors mention vascular injuries, bowel injuries, genitourinary injuries, and port-site metastases (PSMs).^{20,21} Nevertheless, Chi et al²² found that both simple and complex laparoscopic procedures can be performed by a gynecologic oncology service with a low rate of complications. Among the mentioned complications, we consider PSMs rather important in laparoscopic oncology. Also, it should be underlined that PSM is a strong risk factor for peritoneal dissemination.²³ PSMs are associated with poor outcome of patients²⁴ and represent significant patient morbidity and end-of-life care issues.²⁵ All the above-mentioned factors highlight the high actuality of PSMs in today's laparoscopic surgery (Figure 1).

Incidence

The first paper describing the case of developing local tumor metastases after laparoscopy was presented by Döbrönte et al²⁶ in 1978. Following that, numerous data regarding this complication have been published in various surgical specialties. Zivanovic

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Figure 1. Arrow shows tumor metastasis over fascia and subcutaneous tissue. The overlying skin and parietal peritoneum are intact.

et al.²⁷ in their noteworthy study, showed that PSMs were documented in 20 of 1694 patients (1.18%) who underwent laparoscopic procedures for a malignant intra-abdominal condition. In the investigation of Nagarsheth et al,²⁸ 83 patients with endometrial (39), ovarian (29), and cervical (14) cancers were subjected to laparoscopic treatment. The overall incidence of PSMs in gynecologic cancers in that was 2.3%. In the study of Martinez et al,²⁹ 1216 laparoscopic procedures were performed in women with endometrial end uterine cancers, and the incidence of PSM after laparoscopy for cervical and endometrial cancer was 0.43% and 0.33%, respectively.²⁹ Rassweiler et al³⁰ found an incidence of 0.18% in 1098 patients who had undergone laparoscopic procedures for urologic malignancies. Shoup et al³¹ examined the incidence of PSMs for upper gastrointestinal tract malignancies and found that port-site implantation after laparoscopic procedures occurred in 0.79% of 1650 patients. Fleshman et al³² noted an incidence of PSM in 0.9% in 435 patients who underwent laparoscopic-assisted colectomy. Summarizing, Terence et al²⁴ reviewed 17 studies, which included 11,027 cancer patients undergoing laparoscopic surgery or diagnostic laparoscopy, and pointed out that PSM is a rare phenomenon, occurring in less than 2% of patients.

Immune response

Data concerning immune response in the development of PSMs are rather rare. However, among the existing ones, the study of Ost et al³³ is of interest. In their study, mice and the syngenic murine bladder tumor cell line were used. The investigators subjected mice to either CO₂ pneumoperitoneum or midline incision. Peritoneal macrophages were collected. The tumor necrosis factor (TNF)-alpha levels were quantified. As the conclusion, the study showed that in a syngenic murine model, CO₂ pneumoperitoneum causes inhibition of peritoneal macrophage TNF-alpha secretion. Inhibition of peritoneal macrophage TNF-alpha secretion may be considered an adverse event contributing to the development of transitional-cell carcinoma PSM, especially if surgical oncologic principles are violated. Prior to this study, Gupta and Watson³⁴ reviewed the literature on immunological changes following laparoscopy and open surgery from Medline and concluded that despite a few contradictory reports, systemic immunity appears to be better preserved after laparoscopic surgery than after open surgery. However, the local intraperitoneal immune system behaves in a particular way when exposed to carbon dioxide pneumoperitoneum; suppression of intraperitoneal cell-mediated immunity has been demonstrated in a number of studies.³⁴ In addition to the abovementioned studies, Kuhry et al,³⁵ in their review article, argue that although laparoscopic surgery for colorectal malignancies may be associated with higher survival rates and lower recurrence rates because of improved immune function, it has also been related to high incidences of PSMs. Reviews in the literature have now shown that incidences of PSMs are comparable to incidences of wound metastases after open surgery.³⁵ Moreover, in their review article, "Immunological aspects of minimally invasive oncologic surgery," Hegarty and Dasgupta³⁶ summarized that laparoscopy results in better overall preservation of immune function than open surgery. Also, a substantial number of recent studies on the topic of immune response in general and gynecologic surgery were identified from Medline by Holub,³⁷ with a conclusion stating that laparoscopic surgery better preserves the postoperative immunological functions compared with the open approach.

Nevertheless, Ost et al,³³ Kuhry et al,³⁵ Hegarty and Dasgupta,³⁶ and Holub³⁷ (referred by our group), as well as other investigators,^{38–40} share a common opinion that the status of the immune response in laparoscopic procedures and its contribution in future development of the PSMs needs to be further investigated.

Pneumoperitoneum

Some investigations showed stimulation of tumor growth by intra-abdominal pressure.^{40–43} Different pressures and their effect on tumor growth and PSMs in a rat model both *in vivo* and *in vitro* were investigated by Jacobi et al,⁴⁰ who showed that tumor cells incubated with CO₂ at 10 and 15 mmHg revealed a decreased *in vitro* intraperitoneal tumor growth in comparison with pressures at 0 and 5 mmHg. As for the studies *in vivo*, increased tumor growth could be observed at laparoscopy at 5 and 10 mmHg compared with the control group. Increase of subcutaneous tumor growth was observed at laparoscopy at 5, 10, and 15 mmHg compared with controls.⁴⁴ Nevertheless, other data demonstrated the stimulation of intra-abdominal tumor growth caused by high-pressure CO₂ and leading to increased pulmonary metastasis.⁴²

Moreira etal⁴⁵ reported that as a result of insufflating gas' distension of abdomen, a high pneumperitoneal pressure is created, which in turn provokes movement of free peritoneal tumor cells, or may cause sloughing or dissemination of tumor cells from viscera into the peritoneal cavity. An increased blood flow of the anterior abdominal wall caused by intra-abdominal high pressure as a result of pneumoperitoneum can be a possible risk factor of PSMs because of the increased circulation.⁴⁶

Carbon dioxide is the most commonly used gas for insufflations during laparoscopic surgery. It is nonflammable, inexpensive, colorless, readily available, and readily absorbed.⁴⁷ There are several conflicting results regarding intraperitoneal tumor growth as a result of CO₂ pneumoperitoneum. In particular, lingli et al⁴⁸ presented a study where intraoperative peritoneal lavage cytology was performed for 36 patients with colorectal cancer during colorectal laparoscopic surgery and for 45 patients with colorectal cancer during conventional surgery. Cytology was examined twice: immediately after opening of the peritoneal cavity and just prior to closure of the abdomen. Malignant cells were not detected in the CO₂ filtrate gas. The incidence of positive cytology in the lavage of the instruments during laparoscopic surgery was 2.78%. The incidence of positive cytology during laparoscopic surgery was 33.33% in the prelavage and 8.33% in the postlavage. The incidence of positive cytology during conventional surgery was 33.33% in the prelavage and 11.11% in the postlavage. The conclusion was that during colorectal laparoscopic surgery, CO₂ pneumoperitoneum does not affect tumor cell dissemination and seeding.⁴⁸ In this study, laparoscopic techniques used in colorectal cancer surgery were not associated with a greater risk for intraperitoneal dissemination of cancer cells than the conventional technique. Ishida et al⁴² came to a similar conclusion after performing an investigation on rabbits, wherein the presence of a trocar may be a factor contributing to PSM, but CO_2 pneumoperitoneum appears not to be a factor.

Controversial data were presented by Hirabayashi et al,⁴⁹ who performed a study on 15 nude mice, which were injected with human gastric cancer (MKN 45) cells with further determining of the effect of pneumoperitoneum by using a scanning electron microscope to study the effects of how tumor cells disseminate to form PSMs after pneumoperitoneum. They found that pneumoperitoneum immediately results in peeling and destruction of the muscular layer of the abdominal peritoneum, increasing the propensity of tumor cell adhesion at port sites and subsequently healing process occurs, leading to scar formation with the presence of entrapped tumor cells. The conclusion of the study was that free cancer cells appear to attach to the injured port sites immediately after CO₂ pneumoperitoneum, and these are associated with the development of PSM after laparoscopic cancer surgery.⁴⁹

Furthermore, the type of gas has also been shown to influence the rates of PSM with helium insufflations being the least likely compared to argon and nitrogen, which were more likely to be associated with PSM.⁵⁰

Wound contamination

According to some authors, the tumor cell entrapment hypothesis is one of the etiologic development factors of the PSM. The essence of this hypothesis, presented in 1989, is that free cancer cells are capable of implanting on raw tissue surfaces including damaged peritoneal surfaces.⁵¹ The tumor cells' destruction by the normal defense mechanisms could be prevented by fibrinous exudates that cover the raw tissue surfaces including damaged peritoneal surfaces in the postoperative period.

In laparoscopic procedures, the specimen is often extracted through a small wound that can increase tissue trauma, which may play a role in wound implantation.^{52,53} The extraction of tumor through a small port site together with the leakage of CO₂ that occurs may induce movement of free tumor cells that have an increased propensity to implant in the traumatized tissue of the wound.⁵³ During the surgical procedure, ongoing passage and extrication of instruments that are contaminated by tumor material owing to the dissection process, may also explain its occurrence.²⁴

Up to 70% of animal studies revealed tumor cell deposition in extraction wounds.⁵⁴ The survey of Paolucci et al⁵⁵ demonstrated that 55% of PSMs were found at the extraction port. Nevertheless, the same study also showed that an extraction bag was used in 11.5% of the patients who developed metastases at the extraction wound. The fact that PSM can be caused by other etiologic factors is proved in the same study. Although direct wound implantation likely plays a major role, there clearly are other etiologic factors because direct wound implantation does not explain the other 40% of patients who develop metastatic disease at nonextraction port sites. Another common port site where metastatic disease can be found is the operating port. Allardyce et al^{56,57} found more tumor cells at operating ports than at assistants' ports, which suggested that wound implantation was caused by contamination of instruments. The studies show that concentration of the tumor cells in wound washings goes as high as 26% and that the tumor cells are able to recover from the gloves and instruments used during surgery.⁵⁸ Numerous investigators have shown instrument contamination with malignant cells.^{52,56,59,60} Instrument contamination can occur by direct implantation of cancer cells after specimen manipulation. These cells can then contaminate the trocars, leading to PSMs. 54 Frequent changes of instruments may predispose to tumor wound implantation. 52,57

It should be underlined that although conflicting data exist from animal and human studies, a general trend is observed toward systemic immune preservation and peritoneal immune depression during insufflation-based laparoscopy. This altered peritoneal immune response could also be an adverse event contributing to the rare development of PSMs.

Tumor-related factors

There are several postulated causes for developing PSMs; of these, tumor aggressiveness appears to be the most favored. It is a truism of cancer biology that the more aggressive the tumor in terms of grade and stage, the more likely that tumor is to metastasize. Thus, the phenomenon of PSMs might simply reflect the biological aggressiveness of the primary tumor.⁶¹

In their article, "Risk factors contributing to early occurrence of port-site metastases of laparoscopic surgery for malignancy," Wang et al⁶² discovered that the majority of recurrences were in patients with adenocarcinoma cell type, advanced stage (far-advanced disease), and often with diffuse peritoneal carcinomatosis, and, consequently, concluded that PSMs may contribute to the highly aggressive nature of the disease.⁶² We can meet practically the same conclusion in Abu-Rustum et al's⁶³ study—subcutaneous implantation appears to occur in patients with known metastatic disease and is detected in the setting of synchronous advanced intra-abdominal or pelvic metastasis and progression of carcinomatosis.

In spite of this, in a review article that analyzed 31 articles, which included 58 patients, Ramirez et al,⁶⁴ concluded that laparoscopic PSMs are a potential complication of laparoscopy in patients with gynecological malignancies, even in patients with earlystage disease. Meanwhile, Zivanovic et al²⁷ analyzed a prospective database of all patients undergoing transperitoneal laparoscopic procedures for malignant conditions performed by the gynecologic oncology service, in which 2251 patients were involved. The investigators arrived at a conclusion that the rate of port-site tumor implantation after laparoscopic procedures in women with malignant disease is low and almost always occurs in the setting of synchronous, advanced intra-abdominal or distant metastatic disease.¹² Moreover, Rassweiler et al³⁰ performed 1098 laparoscopic procedures for urological malignancies, and concluded that according to their experience the incidence of local recurrence and the risk of PSMs is low and seems to be mainly related to the aggressiveness of the tumor.

Martinez et al²⁹ estimated the incidence of clinically detected PSMs in patients with endometrial and cervical cancer treated at two gynecologic oncology services with extensive experience. During the study period, 1216 laparoscopic procedures for uterine cancer were performed. A total of 921 patients underwent laparoscopic staging for cervical cancer and 295 for endometrial cancer. The overall incidence of PSM in our institutions was 0.4% per procedure (5 patients), and the incidence of PSM after laparoscopy for cervical and endometrial cancer was 0.43% and 0.33%, respectively. Excluding patients with peritoneal carcinomatosis, the rate of portsite recurrence in our series lowered to 0.16%, and the rate of isolated PSMs dropped to 0%. The median time to the development of PSM was 8 months (range 6–48), the median overall survival from diagnosis for all patients was 26 months (range 7-30), and median survival from recurrence was 5 months (range 1-20). They concluded that although PSMs are recognized as a complication of laparoscopy for ovarian cancer, they are a rare complication of laparoscopic staging for endometrial and cervical cancer. The majority of patients with PSM presented with associated synchronous disease.²⁹ Vergote et al⁶⁵ observed a high rate of PSMs after laparoscopy in patients with advanced ovarian carcinoma, as well. In addition to the abovementioned studies, there is a very interesting investigation byNagarsheth et al,²⁸ in which they determined the incidence of PSMs in patients undergoing laparoscopic procedures for gynecologic cancers. The charts of patients treated by laparoscopy for diagnosis, treatment, or staging of gynecologic cancer were studied. No patients without a histological or cytological diagnosis of cancer from the index procedure were included. Fisher's exact test was used for statistical analysis. Eighty-three patients were identified accounting for 87 procedures. The types of cancer treated included endometrial (39), ovarian (29), and cervical (14). Twenty procedures were performed for recurrence of ovarian or peritoneal cancer, and ascites was present in 10 cases. The conclusion was that the overall incidence of PSMs in gynecologic cancers in their study was 2.3%. The risk of PSMs is highest (50%) in patients with recurrence of ovarian or primary peritoneal malignancies undergoing procedures in the presence of ascites.²⁸ Along with this, the study by Obermair et al⁶⁶ is also noteworthy. The given study is a retrospective review of patients presenting with stages 1-4 endometrial cancer, who had a hysterectomy, bilateral salpingo-oophorectomy with or without surgical staging. The surgical intent was total laparoscopic hysterectomy (TLH) in 226 patients (44.3%) and total abdominal hysterectomy (TAH) in 284 patients (55.7%). The conclusion was that the incidence of PSM in early-stage endometrial cancer treated by TLH is low.⁶⁶

Surgical technique

One of the primary reasons for PSM occurrence is the surgical technique used. In the development of PSM, spillage or liberation of cells from the primary tumor has one of the key roles; therefore, handling of tumor during laparoscopy is rather important.^{67–69}

There are several interesting studies performed by Lee et al⁷⁰ at different times. One of the studies involved female mice that underwent crushing of a subcapsular splenic tumor during laparoscopic exploration. The scope of port site involvement in these mice was very high in comparison with those that did not undergo tumor crashing. In the same animal model, the authors found that surgical technique may be a possible factor in port tumor formation. They also noted that PSMs decreased with surgeon experience,⁷¹ and wound recurrence may actually be the result of an unfortunate learning curve.⁷²

Polat et al,⁷³ in their experimental study in rats, referred to the effect of types of resection and manipulation on trocar site contamination after laparoscopic colectomy. The investigators detected and quantified the amount of contamination at the portsite by means of a method utilizing radiolabeled colloid particles following extra- or intracorporeal laporoscopic resection of cecum. Prior to the experimental surgery, they obtained a high concentration of luminal colonic radiotracer activity by per anum application of sulfur colloid molecules labeled with Tc-99m pertechnetate. In three main groups of rats, they either resected a portion of cecum extracorporeally or intracorporeally, or did no resection at all. Each main group was further divided into two subgroups, in which the manipulations were either atraumatic or traumatic. They excised trocar sites as 2-cm doughnuts after completion of the surgical procedure. Gamma camera imaging to quantify the amount of radioactive contamination at trocar sites was used. We detected an overall incidence of contamination in 44% of rats. This rate was 71% and 17% in traumatic and atraumatic subgroups. The resection itself increased the rate and intensity of contamination as well (p = 0.04). The most intensive contamination was detected in the intracorporeal resection with traumatic manipulation subgroup.⁷³ This study proves the significance of tumor manipulation. PSM formation can be a result of tumor extraction without the use of an entrapment sac or by direct dissemination of tumor by contaminated instruments.^{30,67,74,75}

The significance of tumor manipulation during surgery in PSMs is also proved by other studies. In a rat model study, Mathew et al⁷⁶ presented an increased level of metastases due to tumor manipulation in open and laparoscopic surgery. The randomized controlled trial performed by Mutter et al⁷⁷ on rats also pointed out that tumor manipulation is the main factor acting on tumor dissemination in both laparoscopy and laparotomy. In the conclusion, the investigators also highlighted that the laparoscopic surgery had a beneficial effect on local tumor growth compared with laparotomy in the case of tumor manipulation. This beneficial effect of laparoscopic surgery may be related to a better preservation of immune function in the early postoperative period.⁷⁷ Oncological safety of the accurately implemented marcellation of the surgical specimen referred to by many authors should be especially underlined.^{78–81}

In addition, some literature review exists regarding the surgical technique decreasing the risk of PSMs. In particular, Agostini et al⁸² showed in their investigation in rat models that peritoneal closure decreases the risk of PSMs. Schneider et al,⁸³ in their experimental, prospective, randomized, single-blind study, investigated the influence of quality surgery on the incidence of port-site recurrences and concluded that trocar fixation, prevention of gas leaks, rinsing of instruments with povidone—iodine, minilaparotomy protection, rinsing of trocars prior to removal, peritoneal closure, and rinsing of all wounds with povidone—iodine during surgery decrease the risk of PSMs.⁸³

Conclusion

PSMs are seldom encountered. The etiology of PSMs is multifunctional. Recurrence of PSMs is guite rare in endometrial and cervical cancers treated laparoscopically. Nevertheless, a tangible role in PSM incidence is attributed to ovarian cancer, primary peritoneal cancer, presence of ascites and biologically aggressive diseases, surgery-related factors including tumor manipulation and wound contamination. Unquestionable advantages of laparoscopic oncologic surgery should be highlighted. The following key factors may decrease the incidence of PSMs: a surgeon's experience, correct and maximally atraumatical tumor manipulation and marcellation, tumor removal from the vagina, use of an impermeable bag, povidone-iodine irrigation of the laparoscopic instruments, trocar, and port site wounds, and suturing of 10 mm and larger trocar wounds. In our opinion, further investigations of the mechanisms of PSM would contribute to the prevention of this insidious pathology.

References

- Bagley CM, Young RC, Schein PS, Chabner BA, Devita VT. Ovarian carcinoma metastatic to the diaphragm—frequently undiagnosed at laparotomy. *Am J Obstet Gynecol.* 1973;116:397–400.
- Rosenoff SH, Young RC, Anderson T, et al. Peritoneoscopy: a valuable staging tool in ovarian carcinoma. Ann Intern Med. 1975;83:37–41.
- **3.** Rosenoff SH, Devita VT, Hubbard S, Young RC. Peritoneoscopy in the staging and follow-up of ovarian cancer. *Semin Oncol.* 1975;2:223–228.
- Worley M, Slomovitz B, Ramirez P. Complications of laparoscopy in benign and oncologic gynecological surgery. *Rev Obstet Gynecol.* 2009;2:169–175.
- Malzoni M, Tinelli R, Cosentino F, Perone C, Vicario V. Feasibility, morbidity, and safety of total laparoscopic radical hysterectomy with lymphadenectomy: our experience. J Minim Invasive Gynecol. 2007;14:584–590.
- Semaan A, Khoury R, Abdallah R, Mackoul P. Laparoscopic modified radical hysterectomy for early invasive cervical cancer. J Gynecol Surg. 2010;26: 183–187.
- 7. Jarruwale P, Huang KG, Benavides DR, Su H, Lee CL. Nerve-sparing radical hysterectomy in cervical cancer. *GMIT*. 2013;2:42–47.
- Kehoe SM, Ramirez PT, Abu-Rustum NR. Innovative laparoscopic surgery in gynecologic oncology. *Curr Oncol Rep.* 2007;9:472–477.

- Jung JJ, Thain S, He S, Yam KL, Lim YK. Minimally invasive surgery for gynecological cancers: experience of one institution. *GMIT*. 2014;3:73–77.
- Franklin M, Rosenthal D, Dorman J, et al. Prospective comparison of open vs laparoscopic colon surgery for carcinoma: five-year results. *Dis Colon Rectum*. 1996;39:35–46.
- Hoffman G, Baker J, Doxey J, et al. Minimally invasive surgery for colorectal cancer: initial follow-up. Ann Surg. 1996;223:790–798.
- 12. Su H, Huang KG, Yen CF, Ota T, Lee CL. Laparoscopic radical trachelectomy: the choice for conservative surgery in early cervical cancer. *GMIT*. 2013;2:39–41.
- Attwood SE, Hill AD, Murphy PG, Thornton J, Stephens RB. A prospective randomized trial of laparoscopic versus open appendectomy. *Surgery*. 1992;112:S497–S501.
- Barkun JS, Barkun AN, Sampalis JS, et al. Randomised controlled trial of laparoscopi versus mini cholecystectomy. The McGill Gallstone Treatment Group. *Lancet*. 1992;340:1116–1119.
- Southall JC, Lee SW, Allendorf JD, Bessler M, Whelan RL. Colon adenocarcinoma and B-16 melanoma grow larger following laparotomy vs. pneumoperitoneum in a murine model. *Dis Colon Rectum*, 1998;41:564–569.
- Allendorf JDF, Bessler M, Kayton ML, et al. Increased tumor establishment and growth after laparotomy vs laparoscopy in a murine model. Arch Surg. 1995;130:649–653.
- Lee SW, Feingold DL, Carter JJ, et al. Peritoneal macrophage and blood monocyte functions after open and laparoscopic-assisted cecectomy in rats. *Surg Endosc.* 2003;17:1996–2002.
- **18.** Nezhat F. Minimally invasive surgery in gynecologic oncology: laparoscopy versus robotics. *Gynecol Oncol.* 2008;111:29–32.
- Kehoe SM, Abu-Rustum NR. Transperitoneal laparoscopic pelvic and paraaortic lymphadenectomy in gynecologic cancers. *Curr Treat Options Oncol.* 2006;7: 93–101.
- Worley MJ, Slomovitz BM, Ramirez PT. Complications of laparoscopy in benign and oncologic gynecological surgery. *Rev Obstet Gynecol*. 2009;2:S169–S175.
- 21. Munro MG. Laparoscopic access: complications, technologies, and techniques. *Curr Opin Obstet Gynecol.* 2002;14:365–374.
- 22. Chi DS, Abu-Rustum NR, Sonoda Y, et al. Ten-year experience with laparoscopy on a gynecologic oncology service: analysis of risk factors for complications and conversion to laparotomy. *Am J Obstet Gynecol*. 2004;191:1138–1145.
- Chua TC, Yan TD, Morris DL, Sugarbaker PH. Port-site metastasis following laparoscopic surgery. In: Shamsa SA, ed. Advanced Laparoscopy. Croatia: InTech Open Access; 2011:137–149.
- Huang KG, Wang CJ, Chang TC, et al. Management of port-site metastasis after laparoscopic surgery for ovarian cancer. *Am J Obstet Gynecol*. 2003;189:16–21.
 Curet M. Port site metastases. *Am J Surg*. 2004;187:705–712.
- Döbrönte Z, Wittmann T, Karácsony G. Rapid development of malignant metastases in the abdominal wall after laparoscopy. *Endoscopy*. 1978;10:127–130.
- Zivanovic O, Sonoda Y, Diaz JP, et al. The rate of port-site metastases after 2251 laparoscopic procedures in women with underlying malignant disease. *Gynecol Oncol.* 2008;111:431–437.
- Nagarsheth N, Rahaman J, Cohen C, Gretz H, Nezhat F. The incidence of portsite metastases in gynecologic cancers. JSLS. 2004;8:133–139.
- Martinez A, Querleu D, Leblanc E, Narducci F, Ferron G. Low incidence of portsite metastases after laparoscopic staging of uterine cancer. *Gynecol Oncol*. 2010;118:145–150.
- Rassweiler J, Tsivian A, Kumar AVR, et al. Oncological safety of laparoscopic surgery for urological malignancy: experience with more than 1,000 operations. J Urol. 2003;169:2072–2075.
- Shoup M, Brennan MF, Karpeh MS, McMahon RL, Conlon KC. Port site metastasis after diagnostic laparoscopy for upper gastrointestinal tract malignancies: an uncommon entity. *Ann Surg Oncol.* 2002;9:632–636.
- Fleshman J, Sargent DJ, Green E, et al. Laparoscopic colectomy for cancer is not inferior to open surgery based on 5-year data from the COST Study Group Trial. *Ann Surg.* 2007;246:655–664.
- Ost MC, Tan BJ, Lee BR. Urological laparoscopy: basic physiological considerations and immunological consequences. J Urol. 2005;174:1183–1188.
- Gupta A, Watson DI. Effect of laparoscopy on immune function. Br J Surg. 2001;88:1296–1306.
- Kuhry E, Jeekel J, Bonjer HJ. Effect of laparoscopy on the immune system. Semin Laparosc Surg. 2004;11:37–44.
- Hegarty N, Dasgupta P. Immunological aspects of minimally invasive oncologic surgery. Curr Opin Urol. 2008;18:129–133.
- Holub Z. Impact of laparoscopic surgery on immune function. Clin Exp Obstet Gynecol. 2002;29:77–81.
- Highshaw RA, Vakar-Lopez F, Jonasch E. Port-site metastasis: the influence of biology. *Eur Urol*. 2005;47:357–360.
- Wichmann MW, Huttl TP, Winter H. Immunological effects of laparoscopic vs open colorectal surgery: a prospective clinical study. *Arch Surg.* 2005;140: 692–697.
- Sylla P, Kirman I, Whelan RL. Immunological advantages of advanced laparoscopy. Surg Clin North Am. 2005;85:1–18.
- Gutt CN, Kim ZG, Hollander D, et al. CO₂ environment influences the growth of cultured human cancer cells dependent on insufflations pressure. *Surg Endosc.* 2001;15:314–318.
- 42. Ishida H, Murata N, Yokoyama N, et al. The influence of different insufflation pressures during carbon dioxide pneumoperitoneum on the development of pulmonary metastasis in a mouse model. *Surg Endosc.* 2000;14:578–581.

- Wittich PH, Steyerberg EW, Simons SHP, et al. Intraperitoneal tumor growth is influenced by pressure of carbon dioxide pneumoperitoneum. *Surg Endosc*. 2000;14:817–819.
- 44. Jacobi CA, Wenger FA, Ordemann J, et al. Experimental study of the effect of intra abdominal pressure during laparoscopy on tumor growth and port site metastasis. Br J Surg. 1998;85:1419–1422.
- Moreira HJ, Yamaguchi T, Wexner S, et al. Effect of pneumoperitoneal pressure on tumor dissemination and tumor recurrence at port-site and midline incisions. *Am Surg.* 2001;67:369–373.
- Yavuz Y, Rønning K, Lyng O, Grønbech JE, Mårvik R. Effect of carbon dioxide pneumoperitoneum on tissue blood flow in the peritoneum, rectus abdominis, and diaphragm muscles. Surg Endosc. 2003;17:1632–1635.
- Menes T, Spivak H. Laparoscopy: searching for the proper insufflation gas. Surg Endosc. 2000;14:1050–1056.
- Jingli C, Rong C, Rubai X. Influence of colorectal laparoscopic surgery on dissemination and seeding of tumor cells. Surg Endosc. 2006;20:1759–1761.
- Hirabayashi Y, Yamaguchi K, Shiraishi N, et al. Port-site metastasis after CO₂ pneumoperitoneum: role of adhesion molecules and prevention with antiadhesion molecules. *Surg Endosc.* 2004;18:1113–1117.
- Gupta A, Watson DI, Ellis T, Jamieson GG. Tumour implantation following laparoscopy using different insufflation gases. ANZ J Surg. 2002;72:254–257.
- Sugarbaker P, Cunliffe WJ, Belliveau J, et al. Rationale for integrating early postoperative intraperitoneal chemotherapy into the surgical treatment of gastrointestinal cancer. *Semin Oncol.* 1989;16:83–97.
- Neuhaus SJ, Texler M, Hewett PJ, Watson DI. Port-site metastases following laparoscopic surgery. Br J Surg. 1998;85:735–741.
- Tseng LNL, Berends FJ, Wittich PH, et al. Port-site metastases: impact of local tissue trauma and gas leakage. Surg Endosc. 1998;12:1377–1380.
- Bouvy ND, Marquet RL, Jeekel H, Bonjer HJ. Impact of gas(less) laparoscopy and laparotomy on peritoneal tumor growth and abdominal wall metastases. *Ann* Surg. 1996;224:694–701.
- Paolucci V, Schaeff B, Schneider M, Gutt C. Tumor seeding following laparoscopy: international survey. World J Surg. 1999;23:989–997.
- Allardyce R, Morreau P, Bagshaw P. Tumor cell distribution following laparoscopic colectomy in a porcine model. *Dis Colon Rectum*. 1996;39:47–52.
- Allardyce RA, Morreau P, Bagshaw PF. Operative factors affecting tumor cell distribution following laparoscopic colectomy in a porcine model. *Dis Colon Rectum.* 1997;40:939–945.
- Thomas CG. Tumor cell contamination of the surgical wound: experimental and clinical observations. *Ann Surg.* 1961;153:697–705.
- **59.** Hewett PJ, Thomas WM, King G, Eaton M. Intraperitoneal cell movement during abdominal carbon dioxide insufflation and laparoscopy: an in vivo model. *Dis Colon Rectum.* 1996;39:62–66.
- Hewett PJ, Texler ML, Anderson D, et al. In vivo real-time analysis of intraperitoneal radiolabeled tumor cell movement during laparoscopy. *Dis Colon Rectum*. 1999;42:868–876.
- **61.** Sooriakumaran P, Kommu SS, Anderson C, Rane A. Port-site metastasis after laparoscopic surgery: what causes them and what can be done to reduce their incidence? *BJU Int.* 2009;103:1150–1153.
- Wang PH, Yuan CC, Lin G, Ng HC, Chao HT. Risk factors contributing to early occurrence of port-site metastases of laparoscopic surgery for malignancy. *Gynecol Oncol.* 1999;72:38–44.
- 63. Abu-Rustum NR, Rhee EH, Chi DS, Sonoda Y, Gemignani M, Barakat RR. Subcutaneous tumor implantation after laparoscopic procedures in women with malignant disease. *Obstet Gynecol.* 2004;103:S480–S487.
- Ramirez PT, Frumovitz M, Wolf JK, Levenback C. Laparoscopic port-site metastases in patients with gynecological malignancies. Int J Gynecol Cancer. 2004;14:1070–1077.
- 65. Vergote I, Marquett S, Amant F, Berteloot P, Neven P. Port-site metastases after open laparoscopy: a study in 173 patients with advanced ovarian carcinoma. *Int J Gynecol Cancer*. 2005;15:776–779.
- Obermair A, Manolitsas T, Leung Y, Hammond I, McCartney A. Total laparoscopic hysterectomy for endometrial cancer: patterns of recurrence and survival. *Gynecologic Oncology*. 2004:789–793.
- Jones DB, Guo L, Reinhard MK, et al. Impact of pneumoperitoneum on trocar site implantation of colon cancer in hamster model. *Dis Colon Rectum*. 1995;38: 1182–1188.
- Nduka CC, Monson JRT, Menzies-Gow N, Darzi A. Abdominal wall metastases following laparoscopy. Br J Surg. 1994;81:648–652.
- Halpin VJ, Underwood RA, Ye D, et al. Pneumoperitoneum does not influence trocar site implantation during tumor manipulation in a solid tumor model. Surg Endosc. 2005;19:1636–1640.
- Lee SW, Whelan RL, Southall JC, et al. Abdominal wound tumor recurrence after open and laparoscopic assisted splenectomy in a murine model. *Dis Colon Rectum*. 1998;41:824–831.
- Lee SW, Gleason NR, Bessler M, et al. Port site tumor recurrence rates in a murine model of laparoscopic splenectomy decreased with increased experience. Surg Endosc. 2000;14:805–811.
- Zmora O, Gervaz P, Wexner SD. Trocar site recurrence in laparoscopic surgery for colorectal cancer. Myth or real concern? Surg Endosc. 2001;15:788–793.
- 73. Polat AK, Yapici O, Malazgirt Z, Basoglu T. Effect of types of resection and manipulation on trocar site contamination after laparoscopic colectomy: an experimental study in rats with intraluminal radiotracer application. Surg Endosc. 2008;22:1396–1401.

- 74. Whelan RL, Lee SW. Review of investigations regarding the etiology of port site tumor recurrence. J Laparoendosc Adv Surg Tech A. 1999;9:1–16.
- **75.** Iwamura M, Tsumura H, Matsuda D, et al. Port site recurrence of renal cell carcinoma following retroperitoneoscopic radical nephrectomy with manual extraction without using entrapment sac or wound protector. *J Urol.* 2004;171: 1234–1235.
- **76.** Mathew G, Watson DI, Rofe AM, Baigrie CF, Ellis T, Jamieson GG. Wound metastases following laparoscopic and open surgery for abdominal cancer in a rat model. *Br J Surg.* 1996;83:1087–1090.
- Mutter D, Hajri A, Tassetti V, Solis-Caxaj C, Aprahamian M, Marescaux J. Increased tumor growth and spread after laparoscopy vs laparotomy: influence of tumor manipulation in a rat model. *Surg Endosc.* 1999;13:365–370.
- Bishoff JT. Laparoscopic radical nephrectomy: morcellate or leave intact? Definitely morcellate!. *Rev Urol*, 2002;4:34–37.

- **79.** Varkarakis I, Rha K, Hernandez F, et al. Laparoscopic specimen extraction: morcellation. *BJU Int.* 2005;95:S27–S31.
- Meng MV, Miller TR, Cha I. Cytology of morcellated renal specimens: significance in diagnosis and dissemination. J Urol. 2003;169:45–48.
- Shalhav AL, Leibovitch I, Lev R, et al. Is laparoscopic radical nephrectomy with specimen morcellation acceptable cancer surgery? J Endourol. 1998;12: 255–257.
- 82. Agostini A, Robin F, Aggerbeck M, Jaïs JP, Blanc B, Lécuru F. Influence of peritoneal factors on port-site metastases in a xenograft ovarian cancer model. *BJOG*. 2001;108:809–812.
- **83.** Schneider C, Jung A, Reymond MA, et al. Efficacy of surgical measures in preventing port-site recurrences in a porcine model. *Surg Endosc.* 2001;15: 121–125.